

EXPERIMENTAL

Chemicals Phellamurin has been isolated from the leaves of *Phellodendron amurense* by the method of Hasegawa and Shirato¹

Culture Stock culture of *Aspergillus niger* was maintained on agar slants. The growth medium was the modified Czapek-Dox medium with some microelements (FeCl₃ 6H₂O, 20 mg ZnSO₄ 7H₂O, 10 mg MnSO₄ 4H₂O, 3 mg Na₂MoO₄ 2H₂O, 1.5 mg CuSO₄ 5H₂O, 1 mg) glucose 20 g and phellamurin 0.1 g and its pH was adjusted to 4.5 with HCl. The soln. of phellamurin and remaining ingredients were sterilized separately and combined aseptically in the flasks prior to inoculation. 1 l. of the liquid culture medium was inoculated with spores grown on 5 slants and incubated for 4-11 days at 25°.

Isolation 2 l. of liquid medium were filtered and extracted with Et₂O. After removal of Et₂O, the remaining mass was dissolved in EtOH and applied to a column of polyamide. The column was eluted successively with 100 ml each of 0, 20, 40, 60, 80, 100% aq. EtOH. The fractions eluted with 60 and 80% EtOH were concentrated and examined by TLC on silica gel plates with a solvent CHCl₃/EtOAc/HCOOH (5:4:1). Neophellamuretin (*R_f* 0.8) was isolated from silica gel plates with EtOH and recrystallized from EtOH/H₂O.

Neophellamuretin From EtOH, m.p. 189-190° (Found: C, 67.30; H, 5.73; C₂₀H₁₆O₆ requires: C, 67.41; H, 5.61%). *m/e* 356 (25%), Acetate, m.p. 125-126° (from dil. EtOH), $\lambda_{\text{max}}^{\text{sol}}$ 2860, 2833, 1760, 1690, 1610, 1370 (d) cm⁻¹.

Hydrolysis of phellamurin Phellamurin (0.1 g) was hydrolyzed with 0.1 g β -glucosidase. The aglycone neophellamuretin was extracted with Et₂O and recrystallized from EtOH, m.p. 190°. Sugar was determined as glucose with PC.

Acid treatment Neophellamuretin (20 mg) was heated in 5% H₂SO₄ added with small vol. of EtOH at 100° for 3 hr. After evaporation of EtOH, the solution was extracted with Et₂O. Phellamuretin obtained was recrystallized from EtOH, m.p. 221°. Acetate, m.p. 199°. NMR (ppm): 1.33 (6H, s, gem-dimethyl), 1.68 and 2.59 (2H each, *t*, *J* 7 Hz, H-5' and H-4'), 1.99 (3H, s, aliphatic acetyl group), 2.30 and 2.34 (3H each, s, aromatic acetyl group), 5.35 and 5.56 (1H each, *d*, *J* 12 Hz, H-2 and H-3), 6.22 (1H, s, H-6), 7.15 (2H, *d*, *J* 19 Hz, H-3, 5), 7.44 (2H, *d*, *J* 9 Hz, H-2, 6).

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ALKALOIDS OF *DATURA DISCOLOR*

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Key Word Index—*Datura discolor*, Solanaceae, tropane alkaloids, hyoscyne, tigloyl esters, cuscohygrine.

Plant *Datura discolor* Bernh. A species native to the desert regions of south-eastern California and Mexico, sometimes confused with *D. meteloides* DC. which is found in similar locations but may be distinguished from the latter species by the presence of 5 purple flushes in the throat of the corolla and by its pentagonal calyx and characteristic seeds. Taxonomically it accords with Safford's section *Dutra*¹ of the genus. **Previous work** Pharmacognostical description, assay of total alkaloids in morphological parts and characterization of hyoscyne as principal alkaloid². Ontogenetic production of total alkaloids of

¹ SAFFORD, W. E. (1921) *J. Wash. Acad. Sci.* **11**, 173.

² KALIMKARIAN, P. H. and MILLER, O. H. (1957) *J. Am. Pharm. Assoc.* **46**, 393.

Egyptian-grown plants ³ *Plant material* Raised from seeds collected 1959 in Mexico by Dr H S Gentry and kindly supplied by Dr A S Barclay, U S Dept of Agriculture Herbarium specimens of original collection (No 18387) deposited in U S National Museum, Smithsonian Institute, Washington D C and in the Lundell Herbarium, Texas Research Foundation, Renner

Identification of alkaloids Alkaloids obtained by chromatographic fractionation of the Et₂O extract of the dried plant material are recorded below, characterization is indicated as follows P, m p and m m p of picrates, IRP, comparison of IR spectrum of picrate with that of authentic compound, C, co-chromatography (C1–C3, TLC), C1, alumina (Et₂O), C2, alumina (Et₂O–EtOH, 1:1), both visualized with I₂ in CCl₄, C3, silica gel (CHCl₃–Et₂NH, 9:1), visualized with iodoplatinate reagent, C4, paper (light petrol, b p 60–80°C–amyl alcohol–HOAc–H₂O, 1:3:3:3) visualized with modified Dragendorff's reagent

Aerial parts *Total alkaloid*, 0.17% (dry wt) as hyoscyne *Hyoscyne* (principal alkaloid, 0.08%) C2, C3, P, IRP *Apohyoscyne* C1 C2, P, IRP *Norhyoscyne* C2, P, IRP *Hyoscyamine* (0.01%) C2, C3, P, IRP *Meteloidine* C2, C3, P, IRP *Tiopine* C4 ψ -*Tropine* C4 In addition to the above, two weak bases giving alkaloid-positive reactions were detected, one had *R_f* 0.80 (C1) and the other had an *R_f* similar to that of 3-tigloyl-6-acetoxytropane (C1)

Roots *Total alkaloid*, 0.31% (dry wt) as hyoscyne *Hyoscyne* (0.02%) C2, C3, P, IRP *Norhyoscyne* C2, C3, P, IRP *Atropine* (0.01%) C2, C3, P, IRP *Littorine* C2, C3, m p, m m p and IR spectrum of aurichloride ⁴ *Meteloidine* C2, C3, P, IRP *3 α ,6 β -Ditigloyloxytropane* C1, C2, P, IRP *3 α ,6 β -Ditigloyloxytropane-7 β -ol* C1, C2, P, IRP *Cuscohygrine* (principal alkaloid, 0.06%) C1, C2, P, IRP *Tropine* C4 ψ -*Tropine* C4 Traces of bases having *R_f* (C3) similar to those of noratropine and acetoxytropane

The wide spectrum of alkaloids contained in the aerial parts and roots of *D. discolor* is similar to that found in other species of the genus As with *D. metel*, *D. meteloides* and *D. innoxia*, also of the section Dutra, hyoscyne is the predominant alkaloid of the aerial parts (in the same section, *D. leichhardtii* and *D. pruinosa* contain hyoscyamine as principal alkaloid) Cuscohygrine and littorine as constituents of the roots accord with findings⁵ for other members of the genus but cuscohygrine (20% of total alkaloid of the roots, two samples examined) has not been recorded as the main base of other *Datura* roots 3 α -Tigloyloxytropane and tigloidine, two minor components of a number of other species, were not detected

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The cultivation of plants and the extraction and fractionation of alkaloids was carried out as described previously⁶ for *D. pruinosa*

³ SABER, A. H., BALBAA, S. I., EL HOSSARY, G. A. and KARAWYA, M. S. (1970) *Lloydia* **33**, 401

⁴ EVANS, W. C. and MAJOR, V. A. (1968) *J. Chem. Soc.* 2775

⁵ EVANS, W. C., GHANI, A. and WOOLLEY, V. A. (1972) *Phytochemistry* **11**, 2527

⁶ EVANS, W. C. and TREAGUST, P. G. (1973) *Phytochemistry* **12**, 2077